

RESEARCH ARTICLE

Cover cropping impacts on soil water and carbon in dryland cropping system

Hanlu Zhang^{1,2*}, Afshin Ghahramani^{1,2*}, Aram Ali^{3,4}, Andrew Erbacher⁴

1 Centre for Sustainable Agricultural Systems, Institute for Life Sciences and the Environment, University of Southern Queensland, Toowoomba, Qld, Australia, **2** Cooperative Research Centre for High Performance Soils, Callaghan, NSW, Australia, **3** Soil and Water Science Department, College of Agricultural Engineering Sciences, Salahaddin University-Erbil, Erbil, Kurdistan Region, Iraq, **4** Department of Agriculture and Fisheries, Queensland Government, Goondiwindi, Qld, Australia

* hanlu.zhang@usq.edu.au (HZ); afshin.ghahramani@usq.edu.au (AG)



Abstract

Incorporating cover crops into the rotation is a practice applied across many parts of the globe to enhance soil biological activities. In dryland farming, where crop production is highly dependent on rainfall and soil water storage, cover cropping can

OPEN ACCESS,

(20) w*era-a \$•sm_ PLIS ONE

ponem%748

Muhznui

Uni«sty

PAKßTM

P— PLOS ttE oftrawwru:y
ttw revÉW ttwetore, erubk the an of the
cortent of peer revÉwand authw mal,
published artic"s_ tÜry otthis artde
avadabk

O.1371 mal.wne.(286748

Theisan artü

tie terrEot

Canmons Rtnbutkln Limse,whdl

any
saucewe

Mrebut

"lhs

t"

HØh

effects on soil and biological health require further investigation. The of this study was to evaluate the effect of different timing of summer so.um cover crop termination on soil water, total and labile organic carbon, arbuscular mycorrhizal fungi and their mediating effects wheat yield. Through on-farm trial, soil characteristics along with wheat biomass, yield and grain quality were monitored. In comparison with the control (fallow), the early cover crop was the most effective at retaining greater soil water at wheat sowing by 1–4% in 0–45cm soil profile. An increase in water use efficiency, yield and grain protein by 10%, and 5% was observed under early termination. Under the terminated summer cover crop, there was 7% soil water depletion at wheat planting which resulted in 61 % decline in yield. However, late-terminating cover crop achieved the greatest gain in soil total and particulate organic carbon by 17% and 72% arbuscular mycorrhizal fungal growth. A B concentration by 356% and 251 % summer cover crop incorporation resulted in a rapid gain in labile organic carbon, which constituted hotspots for arbuscular mycorrhizal fungi growth, conversely, fungal activities increased labile organic carbon availability. The effect of increased soil water at sowing and over the growing season, organic carbon, and microbial activities contributed to greater yield. The findings that summer cover

cropping with timely termination can have implications in managing soil water at sowing time and enhancing soil water during the season, soil carbon, and facilitating microbial activities

ach

tun &hernrmt and

ot

TteauMurSt.e

while enhancing productivity in the dryland cropping system.

1. Introduction

Soil water is often a limiting factor in Australian crop production regions particularly cropping in the states of New South Wales (NSW) and Queensland (Qld), where stored moisture in the profile at critical emergence, crop establishment and yield [1–5]. However, soil water availability can be managed that changes biological

activities and nutrient availabilities [6,7]. Soil water availability can impair nutrient

availability by affecting nutrient concentration in soil solution and the rate of nutrient transport to the root affecting plant growth and yield. On the other hand, soil water availability

can also affect microbial growth, microbial activities and their physical and chemical processes

that mobilise organic matter via root exudates [9]. There is a range of cropping practices such

as early crop rotations, stubble retention, minimum tillage or no-till and weed control to improve water use efficiency by enhancing capture and preservation of rainfall [10, 11]. Among these, the use of cover crops has been adopted across many parts of the globe to manage both soil water and nutrients but different effects on soil water have been reported for these practices [12–16]. Integrating cover crops into a system can be a method to replace or shorten the fallow duration, which allows longer duration of soil surface coverage before planting the cash crop [17]. Fallow replacement with a cover crop can affect soil water dynamics by regulating soil water evaporation, runoff and drainage [18] and microbial community structure and consequently biological activities [19]. The use of cover crops in cropping systems provides benefits of modifying the soil environment and enhancing soil physical properties through its effect on root-soil interactions, but their impacts on soil physical and biological characteristics were reported to vary in different environments and cropping systems [20–24]. Long-term cover crop practices can lead to changes in soil hydraulic properties, such as soil bulk density, aggregate stability, soil water retention, infiltration, saturated hydraulic conductivity and pore size distributions across the soil profile [17]. Nevertheless, the magnitude of changes can be highly site and management dependent [25–27].

Cover crops can be the source of plant residues from above-ground biomass and root biomass that contributes to the organic matter pool after decomposition, which improves soil hydrology that can potentially increase soil water storage and plant available water [28, 29]. Studies showed that long-term cover cropping contributed to a better developed structure by improving soil pore size distribution and soil hydraulic properties, such as soil water conductivity and retention at the plot scale [30]. However, the impact of cover crop practice on soil water can vary across years and regions that are affected by climate variability and soil types [27, 31, 32]. Under dryland conditions cover crops are more likely to compete for soil water and nutrient with cash crops [15, 31]. Therefore appropriate termination time is crucial for cover crop management to avoid the competition and reduce soil water loss from evapotranspiration, particularly in water-limited regions where soil nutrients and water efficiency are often low [33–35]. Cover crop management has also been reported as an effective practice to improve soil chemical and biological characteristics, such as enhancing nitrogen recycling via reduced nitrate leaching risks, increasing soil organic carbon (OC), and microbial biomass and activities [36–39]. In general, the main drivers of the net change of soil total carbon are the organic matter from plant residues, soil biota metabolisms, and organic amendments which the first two can be supplied or sustained by cropping [40, 41]. Cover crops affect soil organic matter (SOM) and different forms of carbon in soil, i.e., total carbon, organic carbon and different fractions of active or labile organic carbon (LOC) that are often known as the particulate organic carbon (POC) and permanganate oxidisable carbon (POXC or MnOC). Soil OC is the carbon

component of SOM, which can be 58–60% of SOM. POC and POXC are the small and active fractions of the soil TOC. BJOI, and their lability (undergoing breakdowns) has been reported to have a relationship with the biomass of living organisms [43, 44]. LOC fraction as an active soil OC component is mostly derived from fresh organic materials and often correlated with the dynamics of SOM, and is highly sensitive to soil management [45, 46]. Soil with improved SOM management is likely to have higher productivity due to increased LOC [44]. In a semi-arid dryland cropping system, planting cover crops to replace can vitalise soil aggregation through direct addition of SOM, promoting microbial activities, binding of soil particles by roots or fungal hyphae; and alleviation of wet-dry cycles due to evapotranspiration [47].

Improved SOM or soil OC can lead to changes in soil physical characteristics and potentially soil water characteristics [48]. However, the response of plant available water capacity (PAWC) and soil water retention to variation of SOM or soil OC were reported to differ as SOM varies with soil texture [49, 50]. For example, an increase in SOM can decrease evapotranspiration and suppress infiltration and hence increasing soil water retention during cover crop growth stages and improving water efficiency [51]. On the other hand, an increase in SOM may decrease soil water retention for heavy clay soils [49].

In dryland environments, soil microbial communities are considered as another crucial component, which play an important role in coordinating water and nutrient inputs and outputs, which consequently can affect nutrient cycles and hydrological cycles [52]. In particular, arbuscular mycorrhizal fungi (AMF) play a critical role in soil-plant interactions such as stimulating residue decomposition, facilitating plant nutrient and water uptake, and facilitating soil carbon cycling [53, 54]. The interaction between soil water and AMF is often associated with host plants. Soil water availability has a direct impact on plant root lifespan and turnover, consequently, affecting AMF community composition and symbiosis [55, 56]. AMF regulate soil water content through hyphal colonisation and glomalin-related soil proteins (GRSP), which promotes soil aggregation and soil physical structural stability [57, 58]. Meanwhile, GRSP consists of 30–40% carbon and its related compounds were to be beneficial in improving water holding capacity and hydraulic conductivity, and subsequently positively correlated with plant available water content [60]. The presence and decomposition of cover crops can affect the characteristics of the microbial communities, such as variation and structure [61]. Cover cropping, particularly with no-till practice, can not only enhance root colonisation from AMF and possibly shifting AMF community structure during cover cropping season [62], but also enhance early mycorrhizal colonization of the following crop and assist the success of seedling establishment [63, 64]. Cover cropping can be a potential way to improve available carbon, AMF colonisation and population nutrient accessibility [65, 66], and potentially facilitate the symbiotic relationship between AMF and crop roots exchanging water and nutrients for carbon [67].

Overall, cover crop incorporation provides a range of environmental benefits, such as improved soil physical and biological properties, improved soil water and nutrient availability, and reduced soil carbon decline rate [22, 68–71]. Facing the uncertainty of climate variability and increasing food demand, strategic deployment of cover crop practices can be of support for maintaining the function and resilience of agroecosystems [72]. Effectiveness of cover cropping has been reported to vary across many parts of the globe and limited previous works exist on how cover crops affect soil and productivity within a cropping system in Australia, in particular, the state of Queensland [73, 74]. The objectives of this study were to evaluate the effect of summer cover crop practices on 1) soil OC (i.e., TOC, POC and POXC) and soil AMF DNA sequence concentrations at termination time of summer cover crop; 2) soil water across the soil profile (0–150 cm) over the growing and at the sowing time of the following cash crop; and 3) investigate the dependencies between soil OC and soil water at planting, wheat biomass, yield and grain quality.

To address the questions and explore the effects of cover crop on soil health and cash crop yield production over growing in the rainfed agricultural system, on-farm trial was conducted to monitor soil water across the soil profile, soil OC and AMF DNA concentrations, wheat

biomass, yield and grain quality in a wheat cropping system planted after a summer cover crop-terminations of summer cover crops were applied to

manage soil water and carbon- nls aimed to improve our understanding of the plant• soil• water relations in the rainfed cover cropping system in the eastern region of the wheat belt, Where crop production is highly dependent on soil water at planting time and to investigate Whether cover management can influence soil water characteristics and soil carbon accumulation.

2. Material and methods

2.1 Study site

The field experiments were carried out on a farm located north of Goondiwindi in the Southwestern of Queensland State, Australia (Fig 1). The average annual rainfall of the region is approximately 486 mm (summer dominant), with monthly mean temperature ranging from 11.5 to 27.0°C [75]. The study site is a crop growing region in Australia, as it is within the northern part of the grain belt region, where cereal crops (such as wheat, oats, barley, sorghum and maize) are grown in an extremely variable climate [76]. Water supply and storage (soil water storage) are the major limiting factors for dryland grain production in the region [77]. In 2015/2016, the Goondiwindi region's cereal production was the second largest commodity of agricultural production in the region, which accounted for 17% of Goondiwindi Regional Council's total agricultural output in value terms (AUS\$530 million) [78].

The study site had been managed under grain cropping systems by the landowner.

The site experienced long fallow in 2019 due to the drought condition which could potentially alter soil microbial composition and activities (e.g., microbial decomposition of soil organic matter) [79]. In 2020, winter wheat was planted in May and in October, with wheat stubble left standing in the field.

Experimental trials are part of the project the Broadacre Cropping Initiative (BACI) supported by the Queensland Government (Department of Agriculture and Fisheries) and the University of Southern Queensland. All the approvals have been obtained for conducting this research and such as property name and coordinates cannot be disclosed for confidentiality. Our trials were conducted in 2021 with extended summer rainfall (Jan–May) 2% below the 1990–2020 average, which rainfall distribution over three months was greater than average only at the end of summer (sowing time of summer cover crop) but became significantly lower than average three months before planting the winter wheat in late May (Fig 1). Thus, the examined year is a good example of a seasonal condition in an area where winter crop yield is highly dependent on soil water storage. The soil is classified as a vertisol [80] with a high clay content that ranged between 40 and 60%, have shrinking and swelling characteristics in response to the changing soil water content which is related to the changes in interparticle and intraparticle porosity [81]. Fig (C) shows examples of the cracking soil surface at the trial site.

2.2 Trial design

Field trials were conducted during the 2021 summer and winter trial design for the cover crop season was a randomised complete block design [82] under the uniform paddock condition with five replicates per treatment, including fallow treatment as control. Cover crop plots were terminated by spraying at three different stages: early, mid and late stages (Table 1). For the comparison, the control plots remained as fallow as is a common practice during the summer season before planting the cash crop. Therefore, a total number of 20 zero-tilled plots were used for the summer cover crop trial with 5 replicates for each treatment. For the winter season, the trial design was based on a split-plot design [83] which equally

(a)

(b)

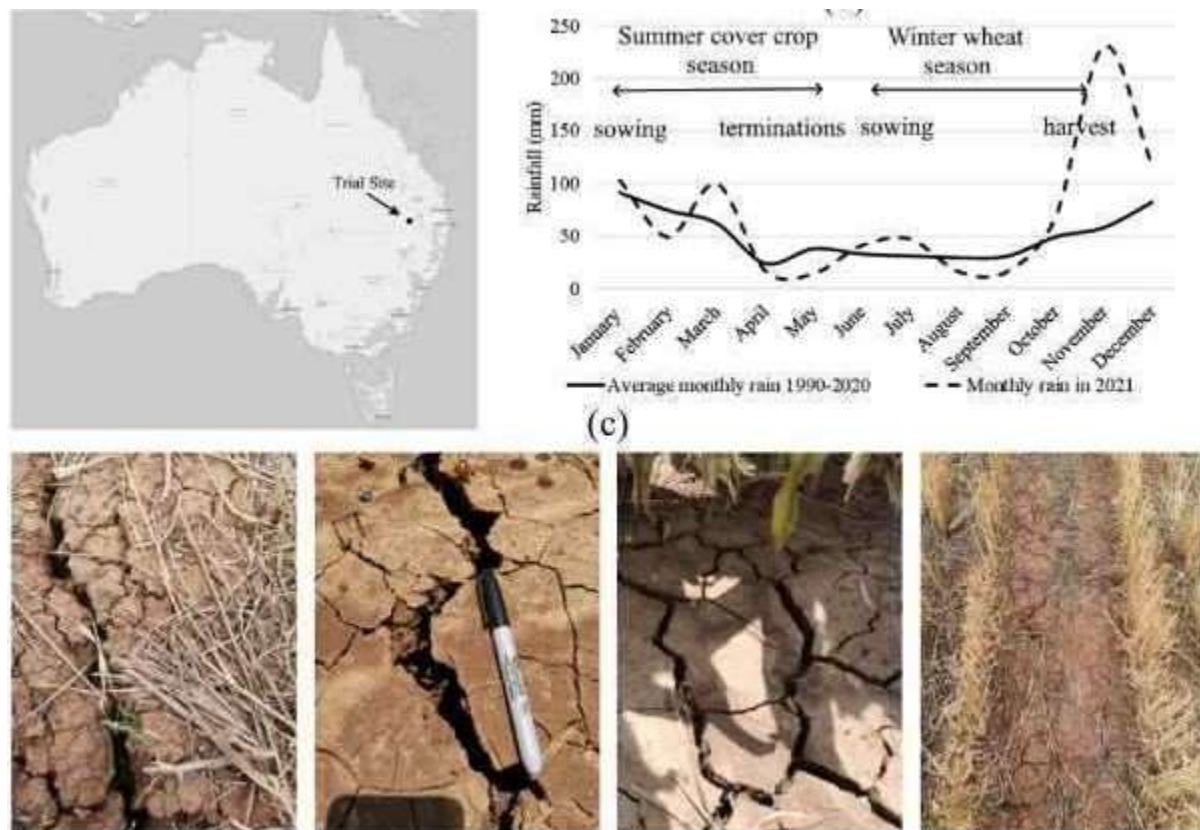


Fig 1. Case study site (a) approximate location of trial site, (b) monthly average rainfall of the season in comparison with the long-term average, (c) soil surface at the trial site (Photos taken by Hanlu Zhang in 2021). Cracking soil surface is a characteristic of the vertosols of the study site.

<https://doi.org/10.1371/journal.pone.0286748.g001>

divided each plot into 2 sub-plots. 'me the subplots Were wheat and fallow (as control)- "Therefore, a total number 20 sub-plots Were planted with Wheat and another 20 plots were as fallow.

2.3 Sampling

Soil from replicate plots of each treatment conducted during the summer cover crop trial and wheat trial in 2021. The sampling included collecting soil cores for measuring soil physical characteristics such as bulk density, particle size distribution, soil water content across the profile, and surface soil samples measuring soil OC and its active fractions and the AMF DNA concentrations (Table 2). The related soil attributes were collected to evaluate the effect of cover crop and its termination management on soil water, OC and labile fractions, and AMF DNA sequence concentrations over the growing seasons as presented in Table 2. These relevant soil attributes were collected for analysis at a system level.

2-3.1 Soil water. Soil profile samples were collected measurement of bulk density, particle size distribution, and gravimetric and volumetric soil water content. Intact soil cores (43 cm diameter by 50 cm) up to 15 cm of soil profile were collected from each treatment plot using a hydraulic soil sampling. Soil cores were cut into 10.0 cm height sections and placed into PVC columns for storage and transportation. Soil cores were preserved in the museum for 48 hours to

Table 1. Components of the farm system at the trial site. Weather records are for the growing season (Jan–Nov 2021).

Weather	Records
Rainfall (mm)	542.8
Mean max temperature (°C)	25.7
Mean min temperature (°C)	12.3
Summer cover crop trial	Trial information
Plot size (m)	18 x 6
Crop cultivar	sorghum (MR Bazley)
Summer control (SC)	Fallow (sprayed and no weeds)
Planting date	15/01/2021
Sowing depth (cm)	3
Row space (cm)	25
Early termination (ET)	2/03/2021 (flag leaf emergence)
Mid termination (MT)	30/03/2021 (soft dough)
Late termination (LT)	21/04/2021 (hard dough)
winter wheat trial	Trial information
Crop cultivar	wheat (Suntop)
Winter control (WC)	Fallow (sprayed and no weeds)
Sowing depth (cm)	30
Row space (cm)	25
Planting date	28/05/2021
Harvest date	27/10/2021
Chemical used	Application rate
Herbicide application	Roundup UltraMax 0.2 ml/ha applied at each termination (Active ingredient 570g/L Glyphosate)
Fertiliser application	Starter Z 25kg/ha on 15/01/2021 and 40kg/ha on 28/05/2021 (Mono ammonium phosphate plus zinc, containing 10% N, 22% P, 2% S and 1% Zn)

<https://doi.org/10.1371/journal.pone.0286748.t001>

laboratory for initial field weight and oven-dried weight through oven dry at 105°C for a minimum of 48 hours to determine the bulk density and gravimetric soil water content. Due to the cracking clay characteristics of the vertosol soils, it is challenging to accurately measure soil water content among various proximal sensors [85]. For this study, neutron moisture meters (NMM) were used to regularly measure point-source soil water in the field. Soil water content measurement using NMM has a better representative value as the measurement sphere is up to a 15 cm radius around the emitted neutron source. In this way, soil cracks are less likely to affect the reading (Fig 2). Soil water monitoring using the NMM approach was based on the physical interaction

of radioactive neutrons with hydrogen atoms, and it had better control of both bulk density and hydrogen atoms during the calibration process [18, 61]. On the basis of the positive relationship between the relative neutron count rate and volumetric soil water, soil water content was estimated from the NMM readings (Fig 2). Prior to the planting of the summer intercrop, a total number of 20 aluminium NMM access tubes were installed at the centre of each plot, which allows taking neutron counts for each soil depth at 15 cm, 35 cm, 45 cm, 55 cm, 75 cm, 105 cm and 135 cm. At the end of the summer trial, NMM access tubes were all removed. The preparation of winter wheat planting—40 tubes were reinstalled after planting and resumed NMM reading measurements for all 40 subplots. NMM readings were taken regularly as part of soil water monitoring during the growing season. NMM readings were

Why measure? What does this represent?	in soil water
physical characteristic of soil represents soil structure and compaction. Can be an indicator of soil health in response to changes in management.	Blake and Hartge 1986 [87]
physical characteristic of soil that drives water holding capacity and flux movement.	Gee and Bauder 1986
ie % soil water on a dry-mass basis; critical to plant growth, nutrient movement and microbial activity	Reynolds 1970 [89]
ie ratio of soil water volume to the volume of soil, can be calculated from measured bulk density multiplied by BD	
included in soil organic matter; C component of organic compounds; An indicator of soil and biology.	Sweeney and Rexroad 1987 [90] Etheridge et al. 1998 [91] Blair et al. 1995 [93]
or partially decomposed plant residue and animal matter with identifiable cell structure. Makes up 2–25% of total soil organic matter; labile OC pool [92]	Cambardella and Elliot 1992
-pool of labile soil OC is defined as the C that can be oxidized by potassium permanganate (KMnO4) [92]	
important microbial communities that regulate plant growth [58] and contribute to soil aggregate formation and stability [57]	Sanders et al. 1995 [95]; Senés-Guerrero and Schürler 2016 [96]

Table 2. Soil attribute tests performed in this study and the purpose of conducting those tests.

Attribute	Why measure? What does this represent?	Method reference
Bulk density (BD)	A physical characteristic of soil represents soil structure and compaction. Can be an indicator of soil health in response to changes in management.	Blake and Hartge 1986 [87]
Soil Particle Distribution	A physical characteristic of soil that drives water holding capacity and flux movement.	Gee and Bauder 1986 [88]
Water (VSW)	ik It i•	
Total Organic Carbon (TOC)	Stored in soil organic matter; C component of organic compounds; An indicator of soil health and biology.	Sweeney and Rexroad 1987 [90]; Etheridge et al. 1998 [91]
Particulate Organic Carbon (POC)	Fresh or partially decomposed plant residue and animal matter with identifiable cell structure. Makes up 2–25% of total soil organic matter; labile OC pool [92]	Blair et al. 1995 [93]
Permanganate-oxidisable Carbon (MnoxC or POXC)	A sub-pool of labile soil OC is defined as the C that can be oxidized by potassium permanganate (KMnO4) [92]	Cambardella and Elliot 1992 [94]
Arbuscular Mycorrhizal Fungi (AMF)	Important microbial communities that regulate plant growth [58] and contribute to soil aggregate formation and stability [57]	Sanders et al. 1995 [95]; Senès-Guerrero and Schüßler 2016 [96]

<https://doi.org/10.1371/journal.pone.0286748.t002>

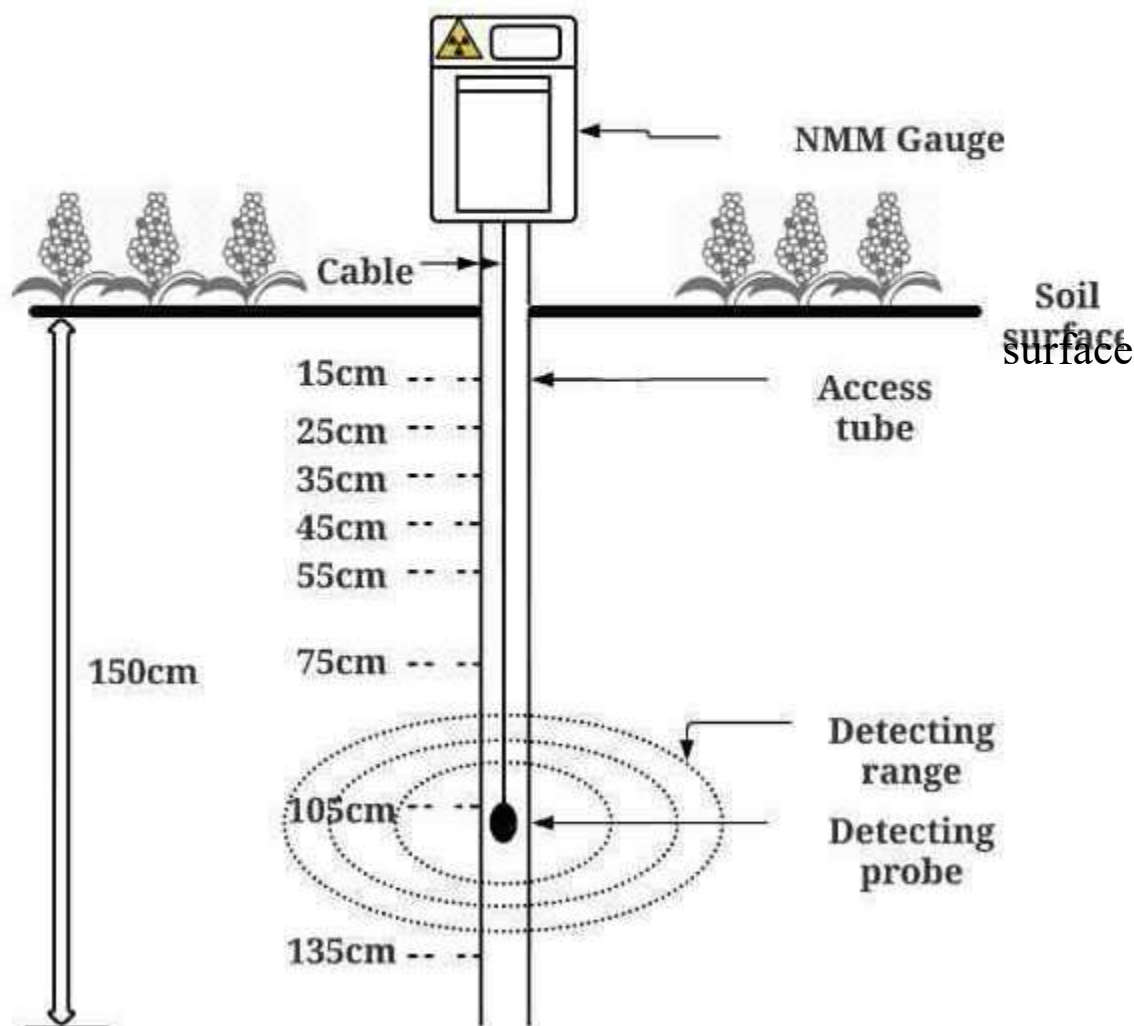


Fig 2. Use of neutron moisture meter probe in the field.

<https://doi.org/10.1371/journal.pone.0286748.g002>

collected by a licenced technician at weeks 4 and 8. Using the dry weight of the soil cores (43 mm diameter by 500 mm length) collected from each depth at the time of neutron probe access tube installation, soil bulk density and gravimetric water content were measured. then converted to volumetric soil water content. The neutron probe was calibrated against gravimetric soil water content, soil texture and bulk density each plot. Soil particle size distribution of the whole soil profile was measured for all plots.

2-3.2 carbon. Top 10 cm soil samples were collected (10 subsamples plot) at the termination time of summer cover crop to monitor soil OC content and its labile fractions POC and POXC (Table 2) to be explored along with soil water content. At the end of the winter trial, soil samples were sampled for OC and POC contents (10 subsamples for subplot). The soil samples were tested by commercial laboratories (Chemistry Centre, Department of Environment and Science, Queensland Government, QLD; the Environmental Analysis Laboratory, South Cross University NSW), to determine total OC content [89, 90], POC [91] and POXC [92]. All samples were air-dried at 40°C and ground to pass through a 2 mm sieve before the 1% instrument used for total OC measurement was TruMac Carman/Nitrogen Determinator (LECO Corporation, St Joseph, MI, USA). The carbon content of the soil samples was determined by analysing the amount of carbon dioxide produced from the combustion of the sample at a high temperature based on the Dumas combustion method [93]. Measurement of POC content was measured by digesting soil samples in Calen (sodium hexametaphosphate) to extract soil fraction >50 μm, then processed for the carbon determination in LECO [94, 95]. Soil POXC measurement used MnO₂ solution (KMnO₄) to react with the soil sample and the POXC content based on the degree of oxidation [96].

2.3.3 arbuscular mycorrhizal fungi (AMF). The soil samples in the top 10 cm were collected from each summer cover crop treatment and their corresponding control plot to test AMF DNA concentrations of different species groups (Table 3). At each plot, 10 subsamples per plot were collected to form one sample (approximately 4 kg sample) that represents the whole plot area. The samples were tested at a commercial lab the South Australian Research and Development Institute (SARDI) laboratory DNA-based characterization and identification of AMF from different phylogenetic taxa groups. In the laboratory, to identify the AMF DNA concentration of each functional group, AMF spores were extracted from the soil samples using Sucrose centrifugation and flotation, followed by polymerase chain reaction (PCR) DNA extraction to form molecular weight 195, 961. The AMF test measured the DNA sequence concentration in each sample and assigned to their phylogenetic taxa using the maximum likelihood method based on near length small ribosomal subunit (SSU) rRNA sequences [97]. The results exhibited the existence of groups A and B (Table 3), these two groups are from the genus *Funneliformis* and *Claroideoglossum* [98]. The functional diversity of AMF such as the function of mycorrhizal symbiosis and its symbiotic efficiency is genotype dependent and can be complex to study the characteristics of species individually [101]. Therefore, for simplicity, the groups A and B were identified based on the DNA sequence, which was used for analysis in this paper, rather than the individual species in each group.

Table 3. AMF species groups and species in each group.

Indicator	Group	Species
Arbuscular mycorrhizal fungi (AMF)	Group A	<i>Funneliformis mosseae</i> , <i>Funneliformis constrictum</i> , <i>Funneliformis coronatum</i> , <i>Funneliformis geosporum</i> , <i>Funneliformis verruculosum</i> , <i>Funneliformis caledonium</i> and <i>Funneliformis fragilistratum</i>
	Group B	<i>Claroideoglossum claroideum</i> and <i>Claroideoglossum etunicatum</i>

<https://doi.org/10.1371/journal.pone.0286748.t003>

(Table 3). AMF DNA sequences in soil and their variation under different summer cover crop treatments were observed to explore whether there were linkages to the changes (e.g., soil treatments Water)-

23.4 Crop. During the summer Cover crop trial, above-ground sorghum biomass was sampled, at the time of each termination using a 0.25 m² quadrat and five random sampling within the plots. The samples were oven-dried at 70°C for 72 hours and then weighed to measure dry biomass weight. During the Winter above-ground wheat biomass from each plot was collected at two different growth stages grain filling and early maturity phenology stages, and collected yield at the harvest. The biomass was oven-dried and weighed using the same procedures explained earlier. Grain samples were analysed by a commercial lab (Leslie Research Centre, Department of Agriculture and Fisheries, Queensland Government) to test grain quality i.e., grain protein, and wheat screenings. A near-infrared transmittance and Dumas combustion (LECO) were applied for the measurement of the nitrogen (N) content to calculate the protein content based on $\text{Protein\%} = \text{N\%} \times 57 \times 100$. The % of the grain samples that pass through a 2mm sieve slot screen was measured to determine grain size.

2.35 Crop water use efficiency. Water use by crop (WU) was estimated as the difference between the sum of in-crop rainfall and the soil water content at times of sowing and harvest. It should be noted that in here evaluation is assumed to be part of the water use efficiency (WUE) was defined as the amount of grain that is produced per unit of water used by the crop, (i.e., $\text{WUE} = \text{Yield/WU}$).

2.4 Statistical analyses

2.4.1 ANOVA. Equality of the variances (homoscedasticity) for the observed variables was assessed using Levene's test. The dataset was then subject to one-way ANOVA with Turkey HSD (honestly significant difference) Post Hoc test to assess the significant impacts of cover crop treatments on soil TOC, POC and POXC, soil water at wheat planting, wheat biomass, grain filling and early maturity, yield, grain protein and screening size. For those attributes that had unequal variances (resulting P-value < 0.05 based on Levene's test), Games-Howell was conducted nonparametric post hoc analysis. Sources of variation were partitioned into between-group factors (treatment). The mean values of these variables were compared under different cover crop treatments with $P < 0.05$ accepted as being significant.

2.4.2 PCA. Kaiser-Meyer-Olkin (KMO) test was used to determine the sampling adequacy of the observed data, with KMO value closer to 1.0 is ideal while values less than 0.5 is considered unacceptable. The KMO value in the acceptable range as it was equal to 0.687. Bartlett's sphericity was also applied to if the observed variables were ideal factor analysis with $P < 0.05$ being accepted as suitable. Then the dataset was subjected to the principal component analysis (PCA) to interpret our multi-dimension observed dataset and assist with exploring the underlying correlations among observed attributes. IBM SPSS Statistics 27.0 Windows was used for the One-way ANOVA and PCA analysis.

3. Results

3.1 Soil organic carbon affected by cover cropping

At the trial site, soil total organic carbon (TOC), POC and POXC contents in topsoils (0–10 cm) increased at each termination time of the summer cover crop. Early, mid and late terminated plots had greater TOC by 7%, 12%, 17%, and POC by 24%, 72% in comparison with the control

plots (Table 4, Fig 3). POXC contents in early, mid and late termination plots were all lower than the control (Fig 3).

Table 4. One-way ANOVA tests showing the significant level of the observed variables means under cover crop treatment in comparison to summer fallow (control). Comparisons without a significant level were considered statistically insignificantly different i.e., $P\text{-value} > 0.05$. Grey shadowed values: a significance $P < 0.05$ was observed.

Observed Items	Dependent Variable	Early Termination	Mid Termination	Late Termination
SOC at cover crop termination	TOC	0.579	0.123	0.016
	POC	0.806	0.744	0.001
	POXC	0.122	0.578	0.992
Soil water at wheat sowing	0–15cm	0.991	0.623	0.026
	15–30cm	0.611	1.000	0.0005
	30–45cm	0.996	0.082	0.002
	45–55cm	0.994	0.705	0.0001
	0–150cm	0.988	0.472	0.0002
Wheat biomass during growing season	Biomass, Grain Filling	0.238	0.936	0.010
	Biomass, Early Maturity	0.732	0.998	0.007
Wheat yield and grain quality at harvest	Yield	0.728	0.755	0.001
	Grain Protein	0.447	0.343	0.064
	Screening size	0.041	0.902	0.917
Soil OC at the end of winter	Fallow TOC	0.833	0.876	0.179
	Fallow POC	0.885	0.546	0.171
	Wheat TOC	0.636	0.294	0.652
	Wheat POC	0.962	0.138	0.361

<https://doi.org/10.1371/journal.pone.0286748.t004>

At the harvest of the winter wheat which was planted following the summer cover crop (Table 1), greater TOC contents were observed in plots that previously had early, mid and late termination, by 7%, 11%, 7%, respectively, and greater POC by 11%, 52%, 38%, in comparison with plots that were under control during summer (Table 4, Fig 3).

3.2 Soil water storage affected by cover cropping

The termination time of the summer cover crop affected the soil water at the sowing time of winter wheat with the greatest soil observed in early termination plots (Fig 3). The soil water content at 15cm–25cm and 35cm were lowest at late termination plots compared to the control (Table 4). The soil water of the whole profile (0–150 cm) at wheat sowing was observed to be in the order of the highest to the lowest: control > early termination > mid termination > late termination (Fig 3). While a % decrease in whole profile soil water was observed for early termination, soil water increased by 15cm, 4% in 25cm and 1% in 35cm, compared to the control (Table 4). Mid terminated plots had lower soil water at 15cm, 35cm and across profile by 10%, and 3%, compared to the control plots, but no difference was observed at 25cm (Table 4). Soil water at 15cm, 30cm and in the whole profile at late terminated plots were lower than the control by 28%, 18 and 7% (Table 4).

Soil water changed over time in all treatments across the soil profile, but the control plots had the least decline and fluctuation during observations (Fig 4). A decline in soil water was observed for all treatments (Fig 4) suggesting water uptake by the plant, and termination prevented further water loss through transpiration and plant usage. At the end of the summer and wheat sowing time, early termination had similar or even greater soil water compared to the control (fallow) and significantly greater than other treatments (Fig 4). As shown in Fig 5 there was no significant rainfall two months

before wheat planting' but early termination was able to store the received rainfall. In comparison with control plots, soil water contents in mid and late terminations plots were both affected by the delayed

ONE

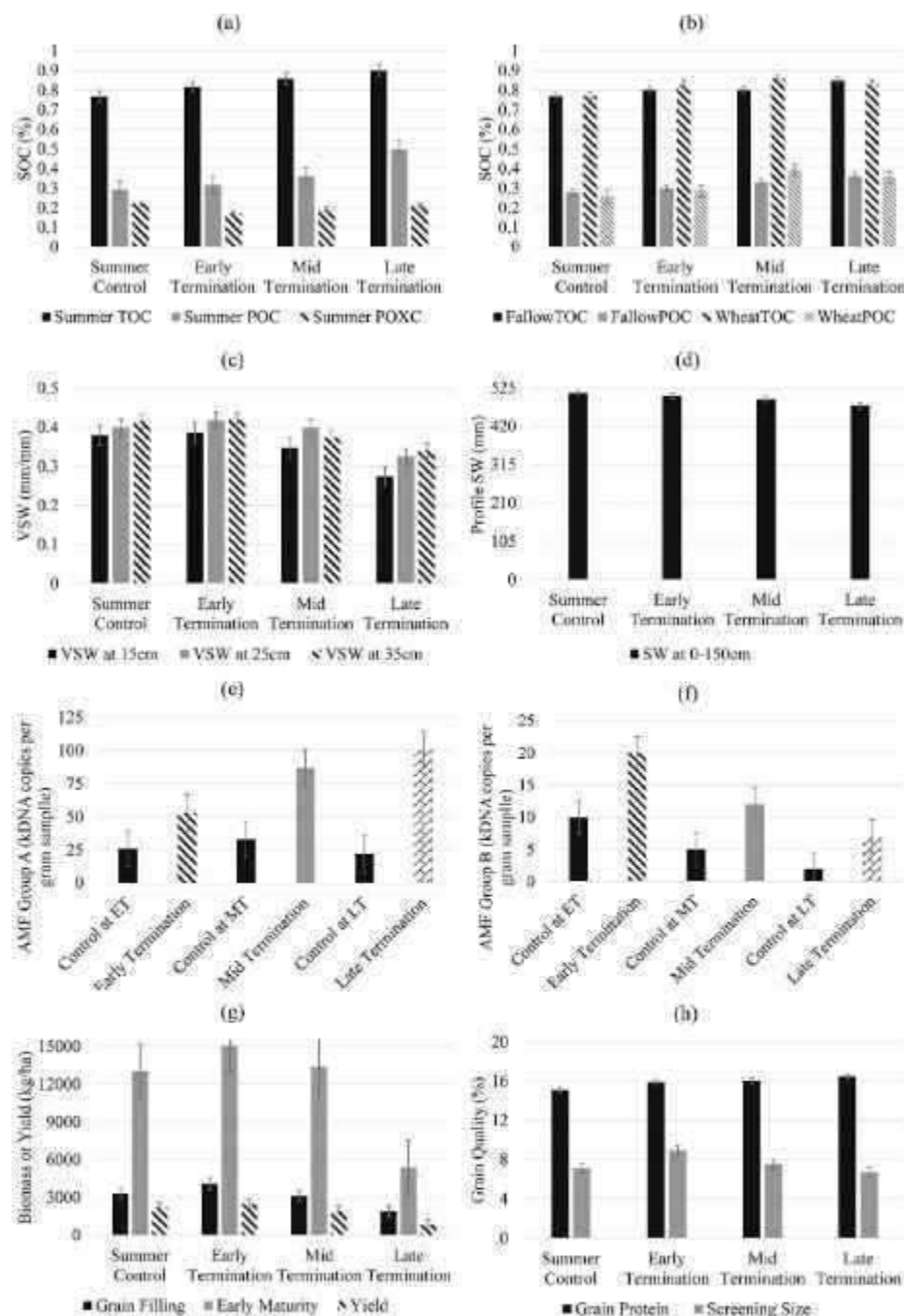


Fig 3. (a) Soil total organic carbon (TOC) and particulate organic carbon (POC) and permanganate oxidizable carbon (POXC) at 10 cm depth at the time of termination of summer cover crop; (b) TOC and POC measured at harvest time of winter wheat; (c) volumetric soil water (VSW) content at 15cm, 25cm and 35cm and (d) whole profile soil water at wheat sowing time; AMF Group A (e) and Group B (f) measured at the time of termination of summer cover crop; (g) wheat above-ground dry biomass measured at grain filling and early maturity phenology stages and yield at harvest; (h) grain quality. SC: Summer control; ET: Early termination; MT: Mid termination; LT: Late termination.

<https://doi.org/10.1371/journal.pone.0286748.g003>

termination, especially in deeper soil layers and the whole profile (Fig 4). There was a 14% decline in the whole profile under late termination compared to the control. Overall, mid and lateterminated cover crop did not show an advantage in preserving greater soil water

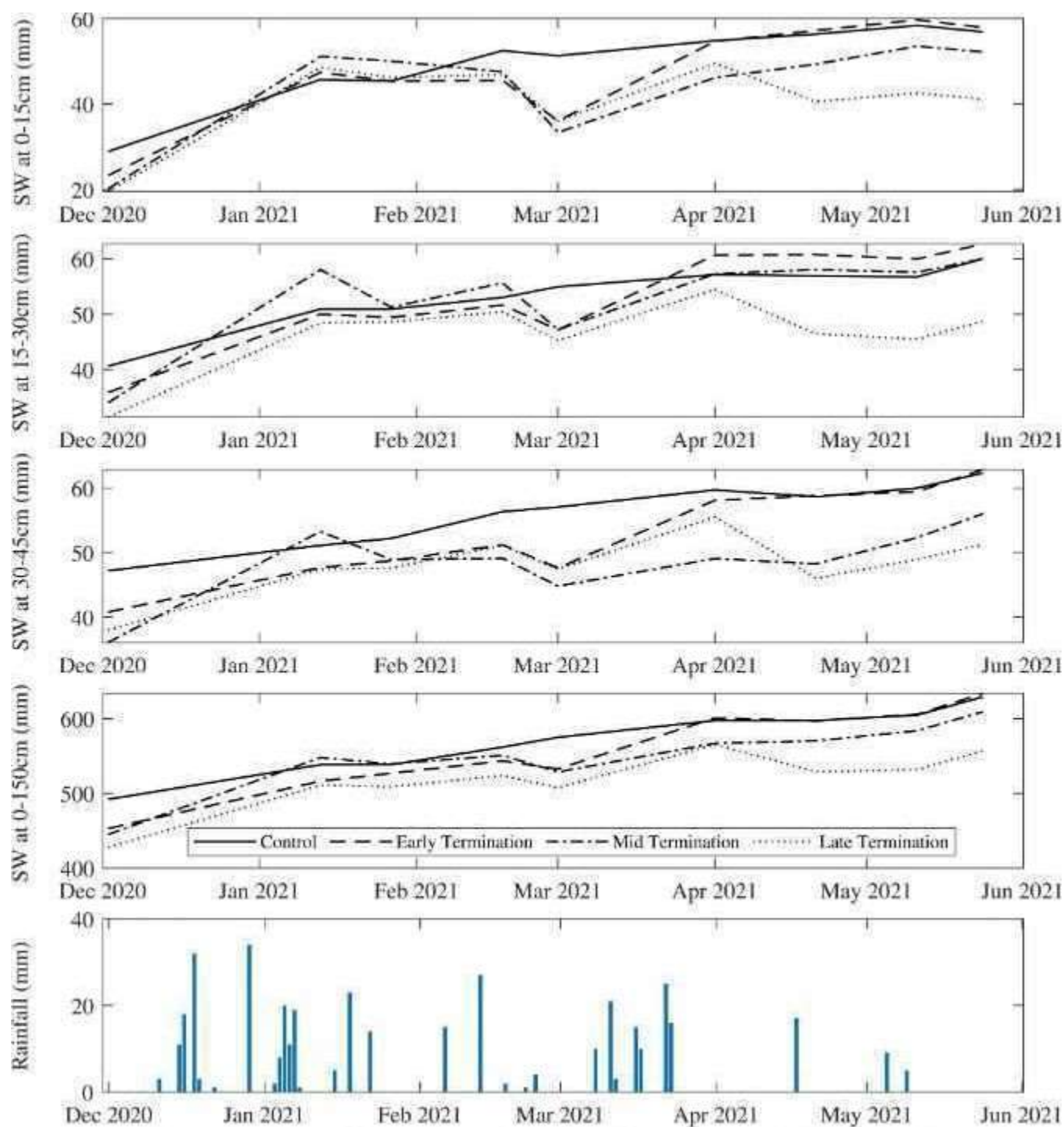


Fig 4. Soil water storage of top layers (0–15cm, 15–30cm and 30–45cm) and whole profile (0–150 cm) during summer cover crop. At the planting time, in the absence of rainfall for 18 days before planting, Early termination had stored higher water in surface soil (0–30cm) while performed same as fallow for all other layers and averaged for the whole profile.

<https://doi.org/10.1371/journal.pone.0286748.g004>

ONE

invadsmsoil water

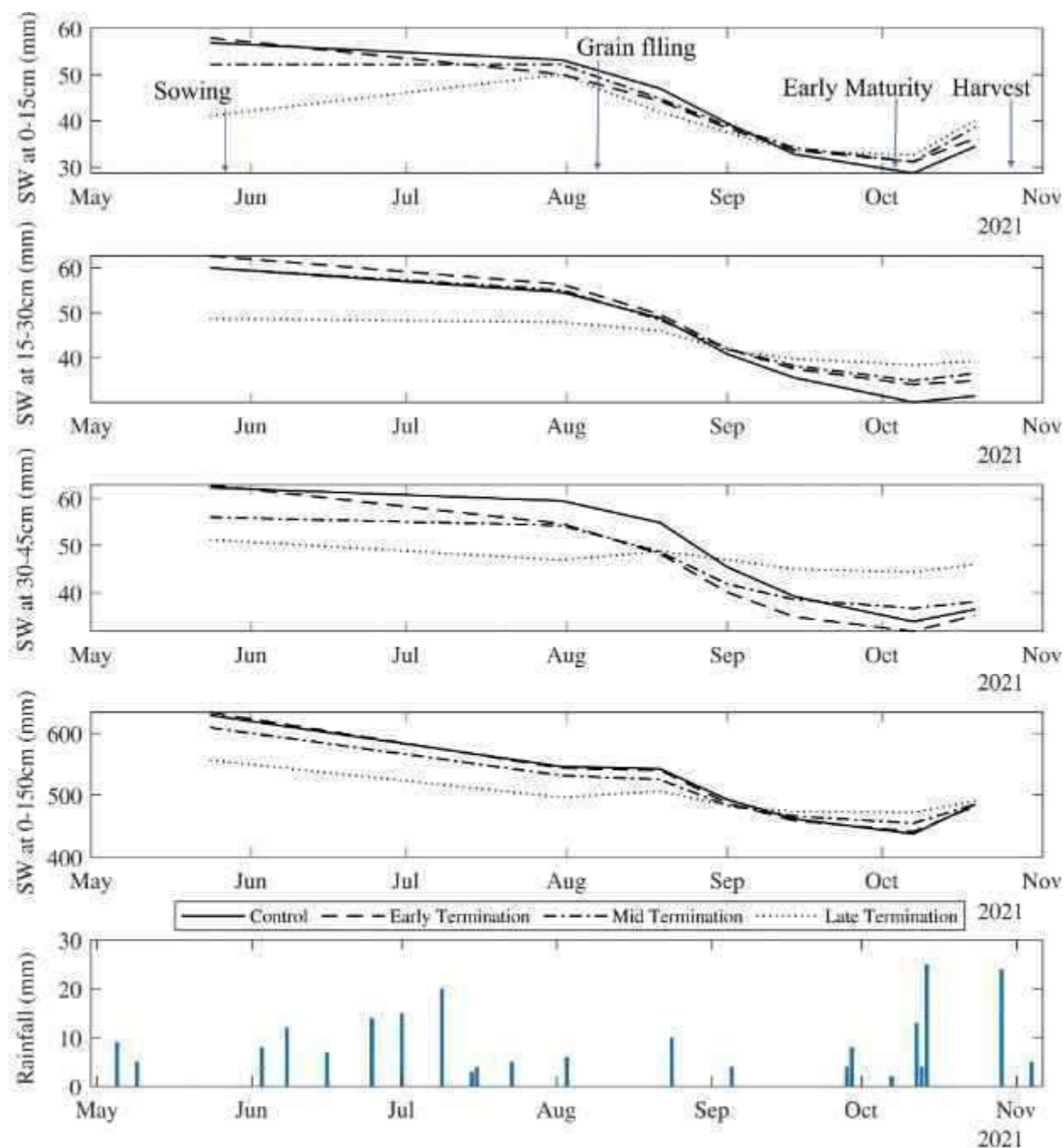


Fig 5. Soil water in topsoil layers (0–15cm, 15–30cm and 30–45cm) and whole profile (0–150cm) during the winter season.

<https://doi.org/10.1371/journal.pone.0286748.g005>

than the control at wheat sowing time, possibly due to little rain received after termination and prior to planting.

3.2.1 Changes in soil water storage over winter cover crop. Soil water in the surface layer (0–15 cm), where wheat was planted on early terminated summer cover crop plots, did layer (0–15 cm), where wheat was planted on early terminated summer cover crop plots did

not significantly over the season when compared to those planted in control plots (Fig 5b). Fallow, early, and mid-terminated plots had almost similar water at the grain filling

stage across the whole soil profile, but water was higher in 15–30 cm depth under early termination (Fig 5). However, soil water was significantly lower at sowing time in plots where Summer Cover Crop was treated with mid and late termination.

At the flowering stage, wheat plots that were planted on early terminated summer cover crop plots had 0.5% less profile soil water compared to the control, followed by 2% less in mid termination and 9% less in late termination. When the wheat crop reached initial grain filling stage, the in profile soil water compared to the control were 0.1% and in wheat plots following early, mid and late terminated cover crops. At the end of grain filling, the whole profile's water differences were lower than the control by 1.8%, 2.1% and 2.5% in wheat plots early, mid, and late termination. Wheat planted on late termination had the lowest soil water content until the grain filling stage compared to the control and the other treatment plots, however, for 0–15 cm layer, soil water was not significantly to other treatments at the flowering stage. By the end of winter (harvest ripe), the highest soil water contents top layers and whole profile were in wheat plots following late termination, followed by mid and then early termination.

3.2-2 Cover crop and water use and Cover crop termination dates. so the amount of WU by crop and in-season rainfall received at each termination treatment was different (Table 5). The early terminated cover crop plots had the opportunity to receive the least in-season rainfall during growth (91 mm), therefore this treatment had less opportunity to water (73.1 mm) compared to the other treatments (Table 5). Contrary, late termination plots used 187.8 mm from 208 mm of rainfall that they received. In winter, all wheat plots received the amount of rainfall 164 mm, but their WU and WUE varied in different plots due to the effect of the previous cover crop treatment in summer (Table 5). Wheat planted on early termination cover crops had the highest WU and WUE compared to the wheat planted on summer control by 10% and 10%. The wheat planted on mid termination plots had 2% greater WUE, though its WU was 14% lower than the wheat planted on summer control plots. The wheat planted on late termination plots had lower WU and WUE than the wheat planted on summer control plots.

3.3 Arbuscular mycorrhizal fungi affected by cover cropping

in control plots. AMY group A DNA sequence concentration (SC) increased from the time of early termination towards the time of mid termination but then declined at late termination (Fig 3). The great AMF Group A SC was observed in late termination plots, followed by mid

Table 5. Water use (WU) by summer cover crop and water use efficiency (WUE) in wheat plots following cover crops.

Variables	Summer Cover Crop Treatment			
	Control (Fallow in summer)	Early Termination	Mid Termination	Late Termination
<i>Summer cover crop season</i>				
Rainfall, planting to termination (mm)	-	91	191	208
WU (mm)	0	73.1±16.9	170.3±7.9	187.8±13.2
Rainfall, termination to planting (mm)	-	131	31	14
<i>Wheat planted following summer cover crop</i>				
Rainfall, planting to harvest (mm)	164	164	164	164
Wheat yield (kg/ha)	2226±219	2500±221	1965±154	870±133
WU (mm)	289.7±13.2	296.8±10.1	250.5±14.3	178.3±10.1
Wheat WUE (kg/ha.mm)	7.7±0.7	8.5±1.0	7.9±0.8	4.8±0.6
ΔWU, Treatment Vs planted on summer control	-	2%	-14%	-38%
ΔWUE, Treatment Vs planted on summer control	-	10%	2%	-37%

<https://doi.org/10.1371/journal.pone.0286748.t005>

ONE

and early termination (Fig 3). AME group B SC control and over cropped plots all decreased the time of early termination towards the time of late termination. Overall, DNA SC of both AME Group A and H, were between the treatment plots and control plots. AMF Group A DNA SC increased by 356% in late termination plots. In mid termination plots and 114% in early termination plots compared to the control. DNA SC of AME Group B in late termination plots was 251% greater than the control. 19% greater mid termination and 1 greater early termination.

3.4 Wheat biomass, yield and grain quality

growth stages, with the first biomass samples collected during the grain tillering stage and the second biomass samples collected during early maturity. Observations that wheat aboveground dry matter from late termination plots was 43% and lower than the control plots, in two biomass sample observations. The biomass in early terminated plots was 23% greater at the grain tillering stage and 16% greater at early maturity compared with control plots. The biomass of mid termination plots was 7% lower at the grain tillering but 3% greater during early maturity compared to the control plots.

The grain yield was highest in early termination plots i.e., higher than control. The yield under mid late termination treatments was 12% and 6% lower compared to the control (Table 6, Fig 3). Among all wheat plots, the highest grain protein content was observed in late termination plots (Fig 3). Grain protein from late termination plots was 9% higher than control, while early and mid termination plots had 5% and 6% higher grain protein (Table 6). Grain screening size early termination plots was 26% higher than control, followed by mid termination (6%), but late termination plots had 6% lower screening size (Table 6).

The one-way ANOVA result showed a significant difference between the variance of cover crop treatment and the control (Table 4). TOC and POC contents measured at termination times of summer cover crop showed a significant difference between late termination and the control (P -value = 0.016 and 0.001). No significant difference in soil POC content was

Table 6. Relative change in soil carbon, soil water, wheat biomass, yield and grain quality affected by summer cover crop compared to the control (fallow).

Plots	Variables	Summer cover crop treatment		
		Early Termination	Mid Termination	Late Termination
Summer cover crop	Δ TOC	7%	12%	17%
	Δ POC	9%	24%	72%
Winter control	Δ TOC	4%	4%	11%
	Δ POC	10%	19%	30%
Winter wheat	Δ TOC	7%	11%	7%
	Δ POC	11%	52%	38%
Crop after summer cover crop				
Wheat, sowing time	Δ VSW, 15cm	2%	-8%	-28%
	Δ VSW, 25cm	4%	0%	-19%
	Δ VSW, 35cm	1%	-10%	-18%
	Δ Profile SW	-1%	-3%	-7%
Wheat, during season and harvest time	Δ Biomass, grain filling	23%	-7%	-43%
	Δ Biomass, early maturity	16%	3%	-59%
	Δ Yield	12%	-12%	-61%
	Δ Grain Protein	5%	6%	9%
	Δ Screening Size	26%	6%	-6%

<https://doi.org/10.1371/journal.pone.0286748.t006>

taund treatments and the control in summer cover Wheat above-ground dry biomass during grain filling and early maturity phenology stages showed significant plots with a history of summer late termination and the control (IL value = 0.01 and $p=0.007$). Wheat yield in plots with a history of late termination was also significantly lower compared to the grain yield from the summer control plots (P-value = 0.001).

No significant was observed in grain protein content between plots with cover crop treatments and the control. Plots with a history of early termination had significantly higher grain screening size (P-value = 0.041) in comparison with control plots (Table 4). Late termination exhibited significantly less soil water across the profile (0–150 cm) at wheat planting compared to the control (P-value 0.026). Overall, the late-terminated cover crop demonstrated advantages preserving soil water, and consequently grain yield, though it was able to significantly increase TOC and POC by termination time.

3.5 Relationships among the variables

PCA results (Fig 6) showed that within the dimension of 1 (of variance). Wheat yield and biomass were closely related to soil water at 15–30 cm and especially in plots with a history of the early terminated cover crop during summer, followed by soil water in 30–45 cm and 0–15 cm. In component 2 (19.4% of variance). PCA revealed an underlying correlation between soil OC contents (TOC, POC and POXC) and clay content (Fig 6). PCA did not exhibit an underlying relationship between grain quality and the other observed variables. Overall, an underlying correlation of OC with clay content and yield with soil water at planting time was observed.

3.6 TOC and Labile OC relationships

Results show that soil POC had a relationship with TOC content, and a greater correlation between soil TOC and TOC was found in summer cover crop plots and wheat plots under

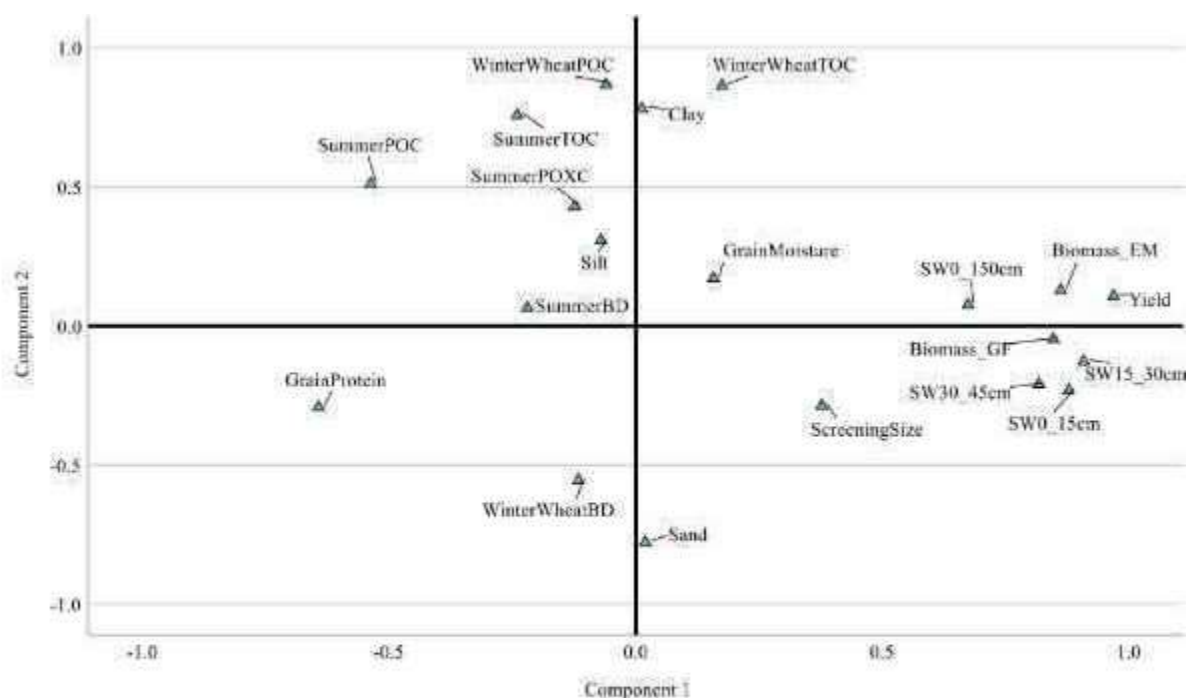


Fig 6. The two-dimensional principal subspace for the observed data. SW: Soil water; BD: Bulk density.

<https://doi.org/10.1371/journal.pone.0286748.g006>

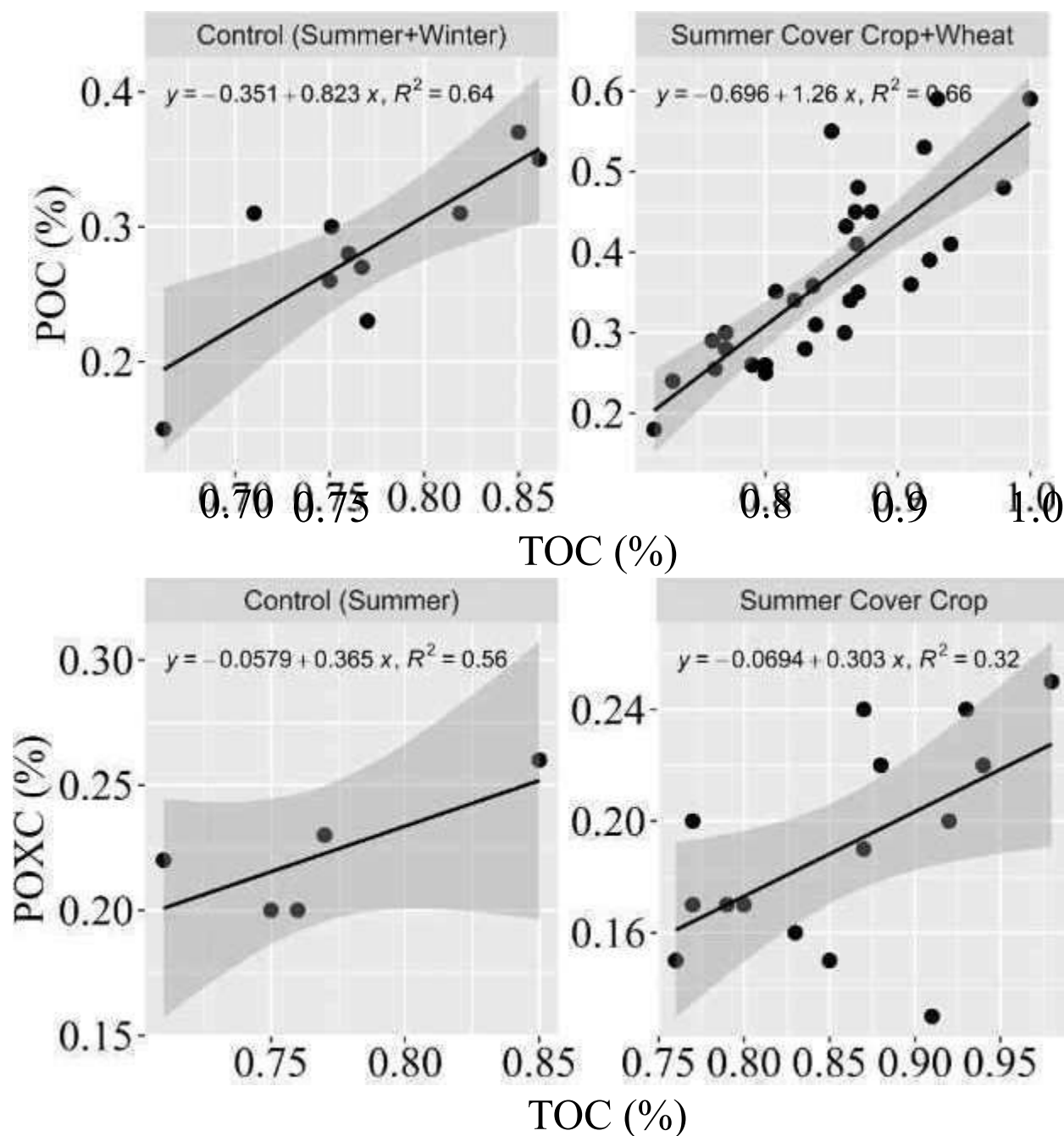


Fig 7. The relationship between total organic carbon and particulate organic carbon (POC) and permanganate oxidizable carbon (POXC). Top left: TOC and POC in summer and winter control plots; Bottom Left: TOC and POXC measured in summer control plots; Top right: TOC and POC in summer and winter cover crop plots; Bottom right: POXC and TOC in summer cover crop plots. The grey shadow area represents a 95% confidence interval.

<https://doi.org/10.1371/journal.pone.0286748.g007>

previous cover crop treatments compared to the control (Fig 7)- Meanwhile, resultsshowed that POXC and TOC in summer had a relationship in control plots. However, the relationship between POXC and TOC was not strongly correlated in cover crop plots (Fig 7). Overall, the relationships between TOC and two labile OC fractions were different in cover crop plots and control plots, with TOC accumulation being more sensitive to the increase of POC esFcia_Ily

under the impact of cover crop management (i.e. the presence of cover crop and how long till termination).

3.7 Yield affected by soil water at wheat planting

Soil at the sowing time of cash crops is critical to seed establishment and biomass production. Soil water at surface layers (0–15 cm, 15–30 cm and 30–45 cm) and soil profile water (0–150 cm) of wheat above-ground dry biomass at both grain filling and early maturity stages (Fig 8). Results also showed that the surface layer (0–15 cm) stored soil water at wheat sowing had a greater effect on yield, compared to the soil water in the whole profile (Fig 9). Wheat planted on early termination plots had the highest yield, while for those planted on late termination, the yield was the lowest. Fig 8 shows that cover cropping practice through the termination dates impacted soil water availability at the planting of winter crop, which the crop's above-ground biomass accumulation and yield.

4. Discussion

Incorporation of cover cropping into a crop-fallow has been practised as a means to manage ground cover, organic matter, stored soil water, quality and health. In this research, the legacy impact of summer cover crops on soil water across soil profile was explored and our results demonstrated the effectiveness of replacing summer cover crop with fallow. ANOVA test showed the significant of treatments stored water, TOC, POC, wheat biomass, yield and grain size.

For the examined season, the early termination of summer cover crop resulted in a 2%, 4% and 1% increase in soil water at planting time at depths of 0–15 cm, and 30–45 cm, respectively, and subsequently led to a 2% increase in wheat compared to the control. Additionally, this treatment increased TOC and POC levels by 7% and respectively (Table 4). The summer cover crop was to enhance soil biology as evidenced by an increase in AMF concentrations in both A and B groups (Fig 3C and 3F). Managing summer cover crop could potentially increase soil water storage during the growing season of the winter cash crop, which could be crucial for sensitive phenology stages (Fig 5). While an underlying correlation was observed between soil water, biomass and yield (Fig 6), the combined changes in soil water and organic carbon resulted in an increased yield (12%) and improved quality, such as an increase in grain protein (5%) in the early-terminated cover crop treatment (Table 4).

4.1 Cover cropping affects soil-water relations

In the current study season, soil water content was by including summer cover crops and by the timing of termination with greater differences observed in the top layers (0–15 cm, 15–30 cm and 30–45 cm) than in the whole soil profile (0–150 cm) (Fig 4). Regardless of the soil water loss due to plant water use, the evidence from trials suggests that the inclusion of cover crops with optimal termination and residue retention creates a beneficial legacy within the soil profile. The early termination was able to retain more soil water compared to the other scenarios (Fig 5) while enhancing chemical and biological indicators i.e., TOC, POC, AMF sequence concentration (Fig 3). This finding provides evidence that cover crop with optimal termination could maximise the soil water storage for a dryland cropping system in the Northern Grain Belt region of Australia. These results apply to our study site for a season with relatively low rainfall but the effect of cover crop management on soil water storage could be different depending on climate variability, soil condition and management [35]. However, this study provides sufficient evidence that cover crop management and its impact on health played a

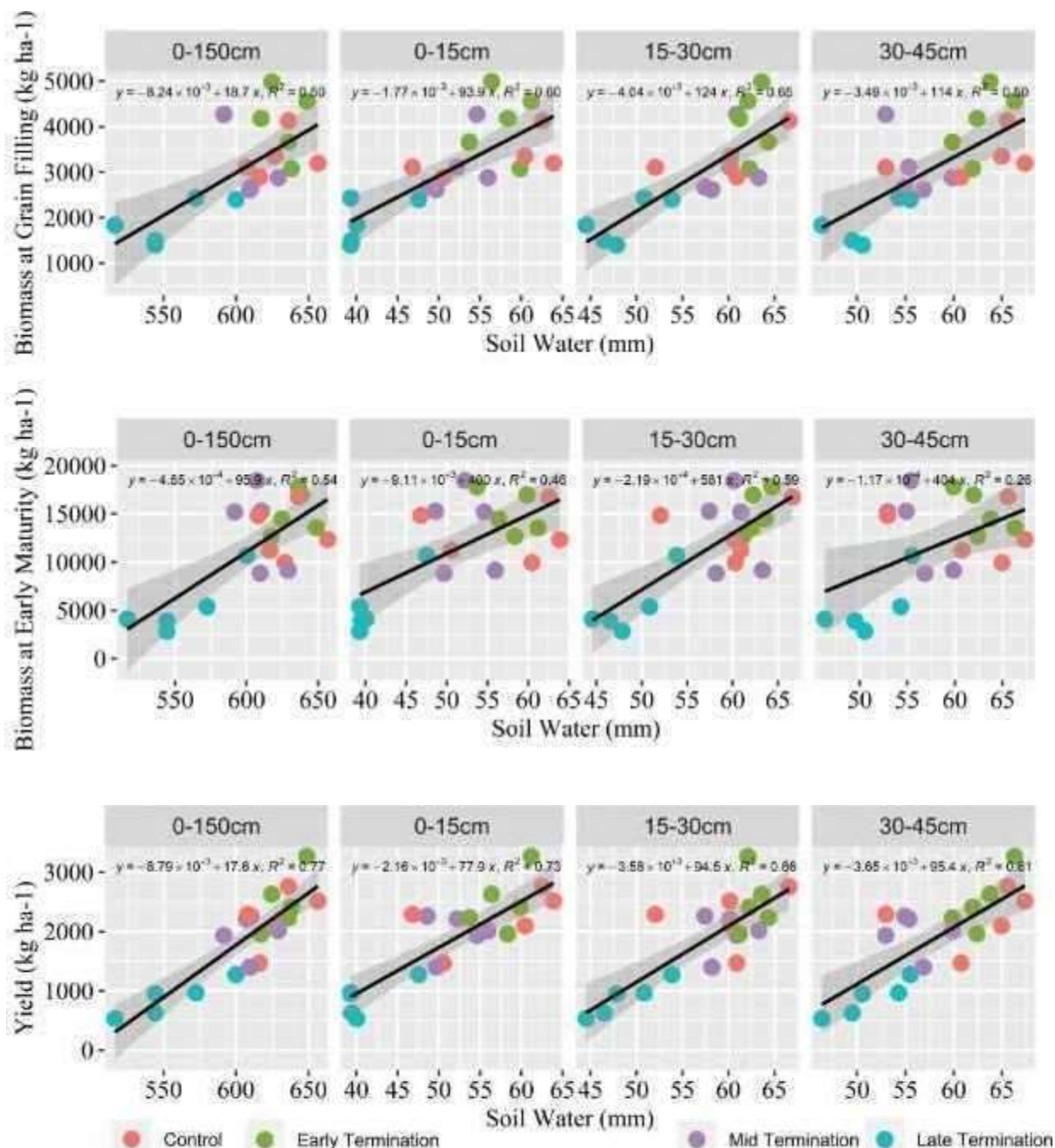


Fig 8. Relationship between soil water at top layers and whole profile at planting time and wheat yield and biomass accumulation at grain filling and early maturity stages. The grey shadow area represents a 95% confidence interval.

<https://doi.org/10.1371/journal.pone.0286748.g008>

substantial role in soil storage and yield. Early termination had similar soil water content at wheat planting time but enhancements in soil health indicators (OC and microbial activity) played a role in the increase in yield and grain quality that was observed. Overall, the evidence from trials suggests that the inclusion of cover crop with optimal termination and residue retention can be effective in retaining soil water while contributing to improve soil biological health (

increased organic matter and microbial activities) and yield production of the following
•e. inter crop.

4.2 Available soil water at planting as a driver of wheat biomass and yield

Winter wheat biomass and yield associated with summer cover crop were attributed to available soil water at sowing time, evidenced by its correlation with wheat above-ground biomass

at the grain filling and early maturity stage (Fig 8). Wheat biomass during grain filling and early maturity response to stored soil water at sowing time was negatively affected by the mid and late termination of cover crop (Fig 8). Due to inadequate soil water at sowing contributed to a reduction/delay in crop establishment and biomass production. Despite the increase in soil OC and microbial activity, the cause of such reduction in wheat biomass and yield was associated with the incorporation of longer summer cover crop treatments i.e., mid and late termination. The summer cover crop was terminated and then left standing, which allowed the soil surface to have ground cover even after the termination. However, mid and late terminated plots had greater above-ground biomass and hence crop residue after termination. A physical barrier of the heavy/dense cover crop residue may lead to an unfavourable/adverse impact on wheat emergence by obstructing light penetration and releasing phytotoxic chemicals from the residue. This phenomenon of crop residue inhibiting plant emergence was also reported in other studies [106, 107].

As shown in Fig 8, Summer cover crop treatments (early termination) could provide soil water similar to summer control at the grain filling stage while facilitating soil biological activities. Soil water availability is critical to wheat root growth and above-ground biomass accumulation, especially for crops during the grain filling stage as wheat has a higher water uptake rate at this stage. Biomass at maturity can affect the final grain production of wheat [108]. Furthermore, dryland wheat growth, grain yield and quality are highly dependent on the amount of soil water storage at the planting, flowering, and grain filling stages [109–112]. Field data at a typical Australian dryland cropping system where the availability of soil water storage and water efficiency are limited [113]. The previous research studies stated that lower soil water availability at planting can lead to a decrease in wheat yield as affected by the incorporation of cover crops. [114, 115], here the finding of this study further highlights the importance of cover crop management and shortening of summer fallow (also called short fallow).

Greater wheat biomass production early termination of cover crop contributed to higher water use efficiency of winter crop, and consequently greater yield production (Table 6). This was due to the combined effect of increased soil water at sowing, soil OC and microbial activities, as discussed in 4.1. With increasing concern about climate change and droughts, the availability of water resources is becoming crucial to dryland cropping systems and system WUE which is often used as a target for soil management [115]. This study indicates that managing soils through proper cover crop management can improve WUE and potentially crop biomass and yield. Cover crop management can be practised for improving productivity via enhanced soil water storage and WUE which can help in the challenges of climate and drought events.

4.3 Cover crop affecting soil organic carbon

Soil to summer cover crop, specifically soil TOC, POC and POXC were different under cover crop treatments and fallow. In summer, the greatest soil TOC at cover crop termination was

Observed in the soil surface layer (0–10 cm) of the late termination plots and was significantly greater than the TOC in control by 17% (Table 4), which could be an outcome of

developed root systems in soils and enhanced biological activities observed by an increase in AMF DNA SC (Table 4). Soil POC content at cover crop termination differed among treatments, With the most significant difference (also the greatest) observed in late termination plots, it was likewise associated With increased soil AMY activities (Table 4). Soil POC constitutes hotspots for microbial activities and has been used as an indicator of soil biological activity. With enhanced soil AMP growth and activities in the late termination plots, AMF was able to facilitate fresh residue decomposition and increase POC availability. Soil POC was considered a indicator for changes in soil quality. 1117–1191 showed that variation in POC can account 69–94% of the changes in TOC due to land use and management. Various other studies have reported similar findings regarding cover crops of species improving soil TOC and POC contents. 1118, 120L Across all treatments, TOC and POC were correlated to each other (Fig 7) suggesting: 1) cover crop management had a consistent effect on improving both TOC and POC availability compared to the control; 2) an increase in POC content contributed to increase in TOC pool.

Different from soil TOC and POC results showed that POXC at cover crop termination was the greatest in the control plots. Followed by late, mid and early plots, but the differences among treatments and control were not significant. This suggests that cover crop management did not significantly affect POXC content over the short term and control plots had a simpler system where POXC was not decomposed/utilized by soil microorganisms as fast as the soils in cover crop plots. 1121–1231. Some studies reported that POXC was sensitive to management practices and could be used as an early indicator of improved soil organic matter management. 1M, 124, 125 but cover crop treatments sometimes can have little on POXC due to low content of soil organic matter. 11261. POC and POXC are the measures of labile organic carbon, the POC method was found to be more sensitive to rapid gain in OC as a result of management or land-use change, while POXC was found to be more sensitive to soil lignin content (a stable component of SOM), instead of rapid gains in OC. 11271. Based on our results, soil POC was more correlated with changes in TOC while less correlation was found between POXC and IOC. (Fig 7). This may suggest POC in our experiments was sensitive to the changes in TOC due to cover crop incorporation. While, as POXC was sensitive to changes in soil lignin compounds which were sourced from surface residue decomposition. Our findings also suggests that: 1) POXC was particularly insensitive to the changes in TOC likely because the trial site had crop residue retained from the previous years and the crop residue has not been decomposed at the time of early or mid termination and hence there was little lignin input in these two treatment plots 2) late termination treatment allowed more time for the residue to decompose (including the wheat stubble from the previous year and fallen litter from the cover crop), and consequently had more lignin input and stimulated POXC accumulation.

Soil OC components measured at the end of the winter season showed that wheat planted on mid termination plots had an advantage in storing more TOC and POC by 1% and 52% compared to the control but disadvantaged yield by compared to the control (Table 4). This was likely a result of a better soil water-microbial environment to handle residue retained in the soil compared to the other plots. Overall, results of this study showed that the short-term cover cropping in summer promoted a rapid gain in soil IOC and POC.

Based on existing studies, the positive relationship between soil OC and crop yield begins to level off when soil OC content reaches approximately 2% [128, 129]. However, no beneficial

correlation between soil OC and yield was observed in trial's soils as OC content was below 1%, which may not be the sole factor driving the grain yield.

4.4 Cover cropping affects arbuscular mycorrhizal fungi groups differently The results showed different DNA sequence concentration of soil AMF Group A and B at termination time, and their response to fallow and cover crop treatments varied (Fig 7). This was consistent with the study of [130] that reported AMF species had different root colonization rates depending on the AMF family (taxonomic variation). The response to AMF colonization differs in host plant species, root growth and the space available for development [131]. Each group's species may have a similar response to the changes in environmental factors such as variation in soil properties and host plant biomass [132, 133] which can occur under cover cropping. The presence or absence of AMF colonization is also related to soil water conditions which in our trials in vertosols, soil fluctuates seasonally to favour or hinder the AMF associations with the host plants [134]. With greater AMF Group A DNA sequence concentration found in late termination soil compared to the control, it was likely because late termination plots had greater sorghum root biomass, which allowed a higher chance for AMF Group A to colonize and establish [135]. The greatest AMF Group B DNA concentration was found in early termination plots compared to the control, and the lowest was found in late termination plots. The decreasing pattern of AMF Group B DNA sequence concentration from the time of early termination towards late termination was possibly related to soil water availability in the rhizosphere [136]. In addition, previous works suggested that intense competition among AMF over root space could lead to competitive dominance in the colonization of AMF species by excluding others [137, 138].

4.3 limitations and recommendations

Overall, this study was subject to potential limitations. Findings of this study were based on the trials within a 1-year window, although with sufficient replications and two examined seasons that had a relatively typical rainfall (Fig 3), a longer term observation might be needed. Our on-farm cropping system research aimed to explore the plant-soil-water relations with implications of summer cover crop practice over the growing of cover crop and cash crop. This has certain significant values to future field studies in the eastern region of the wheat belt as soil water at planting plays a critical role in crop establishment and yield production. The changes of POC (which is a result of short-term management change) did capture the impact of summer cover crop and suggested an improvement of soil quality related to SOM and microbial activities. Therefore, it is recommended that summer cover crop incorporation could help to promote soil health (organic carbon accumulation and microbial activities) through residue retention. This study also showed the importance of the timing of cover crop termination, for its impact on soil water storage. For future studies, it is crucial to consider a number of factors prior to implementing cover crop practices: 1) decision on planting and termination of cover crops should be carefully planned; 2) considering the impacts of cover cropping because it may not necessarily achieve all the benefits (adequate soil water preservation, yield increase, carbon accumulation, microbial health enhancement) that can be achieved by many factors such as condition, growing region, climate, management and investment decisions; 3) considering the potential impact of cover cropping on soil nitrogen retention and their regulation effects on nitrogen cycling processes. For assessing long-term effect of cover cropping practice, it is also recommended to apply validated biophysical modelling to investigate the interactions between soil-crop under the effect of climate variability and management.

5. Conclusion

The implementation of summer cover crop with early termination improved soil biological health and increased soil water content at wheat sowing time, which collectively enhanced wheat yield and grain protein content. This study also highlights the importance of timely termination and residue retention. Cover crop with late termination had some drawbacks such as depleting soil water during the growing and consequently affected soil water availability at wheat sowing time. Although there was evident advantage in soil OC addition and AMP growth under late termination treatment, the loss of soil water at sowing time was detrimental, which led to a significant decline in wheat biomass and yield production. There was a 4% increase in surface soil water at winter wheat sowing time under optimum summer cover crop, but the effects were not proportional to the yield increase, i.e., which suggests that yield increase could be benefited from enhancement in soil health, i.e., soil OC and potentially microbial activities. Overall, summer cover crop practice showed great potential to increase soil health and crop productivity in dryland agricultural systems. Cover crop can be used to manage soil water and soil health, although further research is needed to consider the climate variability and management regime that will maximize the potential and effectiveness of cover crop practice.

Acknowledgments

We would like to express our thanks of gratitude to David Lawrence, David Freebairn, Lukas Van Zwieten, and Terry Rose for their contributions to the discussions. The authors would also like to thank Makhdum Ashrafi, Renier Snyman, James Henderson and Luke Laherty of the Department of Agriculture and Fisheries for their support in conducting field works.

Author Contributions

Conceptualization: Hanlu Zhang, Afshin Ghahramani.

Data curation: Hanlu Zhang, Afshin Ghahramani, Aram Ali, Andrew Erbacher.

Formal analysis: Hanlu Zhang.

Funding acquisition: Ghahramani.

Investigation: Hanlu Zhang.

Methodology: Hanlu Zhang, Ghahramani.

Project administration: Afshin Ghahramani.

Supervision: Afshin Ghahramani.

Writing—original draft: Hanlu Zhang, Afshin Ghahramani.

Writing—review & editing: Hanlu Zhang, Afshin Ghahramani, Aram Ali, Andrew Erbacher.

References

1. Hensley M, Bennie AT, Rensburg LD, Botha JJ. Review of 'plant available water' aspects of water use efficiency under irrigated and dryland conditions. *Water SA*. 2011; 37(5):771–80. <https://doi.org/10.4314/wsa.v37i5.14>
2. Dreccer MF, Fainges J, Whish J, Ogbonnaya FC, Sadras VO. Comparison of sensitive stages of wheat, barley, canola, chickpea and field pea to temperature and water stress across Australia. *Agr Forest Meteorol*. 2018; 248:275–94.
3. Asoodar MA, Bakhshandeh AM, Afrasiabi H, Shafeinia A. Effects of press wheel weight and soil moisture at sowing on grain yield. *J Agron*. 2006; 5(20):278–83.
4. Bell L, Kirkegaard J, Whish J, Swan T, Dunn M, Brooke G, et al. Managing crop differences in soil water extraction and legacy impacts within a farming system. *GRDC Update*. 2021.
5. Verburg K, Thomas M, Cocks B, Austin J, Glover M, Stockmann U, et al. Using existing soil and landscape data sources to estimate plant available water capacity (PAWC) for decision-making and crop

- GFWC; AV 4. AvaA& fran:
•at-al-CO
- ark C , LM, 9øwart A, , M. Eltoas ot past and t*xna— Mid Bd Fend
7. Varp G/ S, J, untotto C, G, Basanta M, Lovera E, al. Fū and q..ahty to J MBobd
 8. P, Z NutriM1t "s. P, t" Lonchrr PtEs; 2012 p.
'C. Wøny A, Blazgwez SJ, Bru D, Rzn•ard N, Brou• MC, al mattMs_ otrpO•
on Etivny rewatt.ögof a systun_ ISME J m18; 12
ICE O. 03Ws41 • 396-0iB007WzPWD•_ 29476139 BM, Hunt JR, A
KJ B, Swan T Gowæd ot al so'
00548 PMID_ 32499799
JR, C, TM, Vo.•tvk K, Cra" S, AM_
ard impacts on winter crop yiej sod wat% arxf N bw rainfall a
of Crop Pagure g;
ES CHTsner TE, Pott« PM Baku JM. ry•e cova nirato, soa
J_ 20t 1;Mar;
Harum S'. NV. crW,
 14. Gabrid Garcia-Gonzalez I, M, Ma D, C
Cavor to water cugent_ —ma
 15. ot covoraop rnixturæ in swd maize cm•or-cn_» ronUm l:
md J Lhn Erv_ 2017;
 16. Fay D, Apria-ti R, Yuwt DYP, Chu D, Famw Wver crops and dymne
in a AM EmGy•st Envirm_.m; htW,'/tmeo-rcV10 to
 17. H, JD, A), Tatarko J, Stuvor TW a sernarUj Efta:ts on .
77
 18. KkBo NL Currie ('E crop ASABE m, 52
103-10.
C, M, MM. The cover
AMF commntnity in so/ md nuns of mcßzo after a crop
Tow Envmm 2019, ogs PMID•_ XJ743976
Yang W. G, Y, J, O P. of covet on rainfM
patterns. X
 - 21, R, M, C, , D, ot al. Cover
Agr
 22. Sumn LM, AK. JD, KL
dryiand Agr Ecosyst Envirm_ am; 32B: 07852
S. Ytamv• V, AT, Dbmm K cowl
Fidd capacity, wemg point, sod-wator
ny, and infinrabm. TransASABE _
 24. CM, W, 11B, Bswas A a
Agb*lturo_ 2021;Mar•, t
M, S". RP, em•er 2019:

H, SJ_

SSSAJ_QC); 84

27. Garba II, LW, WiAarns A. Cover legacy noæts and on "eus a Agrm
2CP2; to 1 æi3-022.(M600
TC, Ж. jM. ctNer пт;
M, J Soe Water Omserv, C, V“navo C, H, E“et1 a
Banh± B, A, BЮnBraП E, cгсф кт атк1 Ы, LX H, FNor C, ВИТ“ B, ап шме rn*ze cumvatkm southern Hoe
Araya SN, JP. де BA,
M, TA and so• waler Soa.
31. PW, V@W. 1 * 3
к—er S , F , ЫВлагШ Sout' Tex“ ШуИ'иJ J
M, pt_os CXЮ, A. tay exacertAte
2014; Сивег•_д)22; 1 ;
M. dategs a
- 34, Л, М_ tnro ta“ow M
ида ам Еш J Чоп. 2011 ; 34(3): 33A3_
Кбуш H, JE, p, E. Covu tnR
a'ways
water u:mlent ю райMaИ шц1 Ay Water Map•
• _ 2020: 231 :1оьддд.
EE, Q Сћепмти P. ТИО uw ot сомот cго to roduco nBraIe
Te ам Ыа'ж»се and ser"e€ 'NRA; 2012 21 L
Ava"aNe
Norhs CE, Cuueves KA. АйеПю'Юе syMems_• a
37. rov•, FESE
И. MG, Dk T. GiatunBM S, Л, BayerC_ а ще•Л•
- ТВОИТТ»Е and &vorsity to Олег са» Igdw•jos_ PLoS 20; 15
PMID: :П36ДЕ0'
- 40, Т*ЮдJE, БЮ. Grat»r EK B“мтаг Тю аЫи'Шгхе,
БЮ'a_ : Lohmarm J , Jo—h S, odaors. Юг штмЧгюппиНа' mut.nmт:
ам 'прИппшитАм1_ ИпЮп: TayЮг& 201S_p. 27—0
Нег«х PE, DA, JM, (Пегпшт SN Nota as си
т: CA, •cw_ sys1«ns CAC p.
42. MSA M O, EI P, M д- art
BmzArch 2008; 51204,
We• HA, WOff F_ 0' от— mawr фПйу ам т: WM
ЯН, иMог& *0.Awe_ CHC Pr—; ЖЮ4, p. 143.
- 44, Uras ST. Cao a Iest to ю
Agroo J_ X 12;Jи';
Soum GP , CC. OM. between frMkrts

- otNy Ay Епч'оп. B;
268%-23_
Wang CH, Wu III АПМy MS, PM'sm D, 2,
soii systams_ SB Agrk: 2016', 73:53542
46. Rj. mattor as Mcator 0' фaBIy Zeakmd_ so• ba at;
47. DenM K. SixJ. H, Frey SD, t>twelon wn:kx H, of cydes шт
BIO BIOB-IOT_ dy'nn«
Murphy BW_ «—с m(ter m:• Res_ 2015;
49. %wts WJ, Pж*цвкy YA, & .•aEr MM, H. Effet1
03; 1 III
MTamTy B, KIB•aПшy AB. 2018:
51. L\ng Y, Hиагщ H, l_ ZQ, WH. E— 0' Еш JSN
шт
mdsn•re dynu-r— 477: Тгмв РнНЮaBIв [M; T 4 ., p. 481—4.
SB, J %robal in .
2012; PMIO: 22772903
ME turW: h•rways tor aruf rurioNs arki VB.
54. Wai Vosâtka M, Ca B, Dom , Lu C, Xu J, rdoot atxnAAar my-corrNza turO
ot a SSSAL 9', t —7-
A, M, D, S, E, Imnnidos 1M. Gerotypo am' so'
ot AMP at st—.
Ecd_ 150t0344a
Wu Y, C, Li J. G_ ot mytM'tuzal
water coru'tk»m_ Pkmt 4644 t): 441
57. Wg MC, SF. Ev•inør VT. Tho rob myconhizal
tkxr øftocts of We am;
N. C, MA, S. Khm , M. etal. ot attn=uhr in gAant growth in abiotic Frmt
PM Sci X)t9; to: https•Ætnc0tosmNjk_2019_01(N PMD: 31608075
S, R, Vanm A, Smrma AK wit'
Giga*Ma rmrgarita PtmtPa1hoL the phr« in viro_ J
alt 7:
tho pm noisturo groMh t B;
g_c030t PMID•_ 24563924
61. Kin N, MC , Guan K. Vilhmil cov« amont
So" 142: 10701_ TM, LE, Loüwr M,
TR.
'tuzas_ a ot crcv J ECOL 17; 17W93.
Moru EM, Avkj •nana C, TurriniA, G, BårtAi P. Fust a mvol
on rryccntüzal h_.ngi md L>v_
I. M. C. activay

- cover a sunmwor Si:A 2016:
- EE. WU*hgs K, McDmOI MD, GP. GraMy AS. Covercrop to a no-W torwrgy
crc»ging systan_ GCB 2017 9(7): 12S2• ST, ME. ME. «atms pr«isctbn "th fgwor
nputs. Agr Ecosyst Envüon. B:
67. Augé RM. Water symÄ
68. AD, Kaspar TC, Archam_As SV, DB, Sauer TJ, Parkin 1B, Sod with tho Ong-tom of a
rye covet crop. Water
H, Mkta WA, Presky DR, Ciaasson MM _ ot cowl nou f« '*wsical
prowrtios_ SSSAJ _ 2011 ;
70. SI , NV. Otteas
Envfon Sci. 201 S: 29• _ 134
- 71, V , J , J msan LS, K,
agotTAü: N , P S Agr Emsyst
Environ.
72. HunW MC, Snot' ME. LW , OA. for 2017; I;
U GD , Hayas , W. Satüal GA, Ihar BSIThw tho pecajctivity of sys, terns P-
ßu_re Sci 20t4;7; t OMB-ttxji.
74. ITuup-Kristmsa• K. J. F-bot tho sy40ms Arm 6; t 10_
rncw122 PMID:
75. Bureau camato o' A" Av— 'run: h ttpj/www tnn_gov auemalß'daw.
Gls HdzwMh DP, Stono R. Tho vaun ot s" to in a with valiaåhty_ AugJ 19%;

77. Оагц УР. PW, ИС, W, ОаИ' НС, III, at aL Suat*
'..ат%щ т AwuahSs гимТш-п •ms: Н опы'гмип«н_ res_ 201 Ют
Б; 152:115-23.
Аимтюп Вигши о! \Тие 0' СкттхТЮд ABS_ ABS cal_ по, 16
НВ.О; Тб, Avant* tom:
79. Рд С, КЛ 3, i_i' У, Н' Х, ecosysErrs_ ESRv. фтиТД
2021; 214:
GA. ГПарМ1 GW. Frmbaim OM, III.
дг&п ot40 years o! Aust J EvAgr. ОП;
47Ш) вв7-вв. 16:
Н. The Austr*nsNBsMabcm, МЮ): СЫРЮ
СЕ, LP, Ихрп о! Мм
Адгоп. ЯМ; 6720-37Б_
Сие••« AG, [Ж.
Ал теу К SMB;
В . , ау, агв1
- (—е GR. КН, Ии: КИТ д, от 1
Апи1сал St"otyot
G G J B. Кыш А, «Мог,, Метив og PT 1 Phу—•
М: SodSbu•ce 38341.
SG_ The gravi.'Wtrt
щобЮты J Ну•drcA.
НА РК LECO FP•228 •пШ.п&юпптаюг• АОАС
Иа.ум J А— Т Иип_ 70: 1 (РВ-Т_ АЛО:
91, HD, GM, ЕН_ А ситра"оп пйтшт апа [jwnas
сотБиМЮп meНух1—es (Уу;о CNS 2(Т) шт 1016'
Агт-п
S0377.B401
Нимч, Д, Впит, Ел— Г, Впл_т S_ 'denMy•jg tofig«pdnt 0' амо
cattxm ад а moasum сатЖт ЕошМ Еw j SN 2tP1 Дие 72(4):
GJ, Letroy Ю, 1_БЮ Sod catN» fm:bons там 0' а сшЖт Мех
ayttAtural systM'& J Адге 1—
СА, Ebott ЕГ. ПарЖ:1Мо
sequeoce_ SSSAJ_ 12560:777-0.
[Н, М М, Grtu»о К, Т. ОнбЛсалЮп 0' 'itxso•М ОНА
апогц шия spots 0' sttbs
C, А. Мew charx1«imlbn and «афиы:иИг
h_mgi Е, (МЕЧ.
101—23
97. Уоопи-в, Вгептш JM Смыт ам
.-,aues_ сопитит S«
ИА, U , ази1 гпаш,,
2 and Ма&мм-• (KM): Эгтд-ег: 1!B3_

- д, W*or C. ТИО Gkkneromycota: a new ала —ra Пю Ноу“ Гип:
201 Ы47Ы2 ве. •-кшу ш, вкю«ю се Хххw «вгйА to _
101. Е-вшптют М, Tay R T, EBstramj M. Func l *ma' тусопЖт—Пю
FOV ЕссИ, 2010;
- 102 OS, RE, wam
Agre%l _ I
Fremh RJ, JE Water dfeigœy whoat a M0'Mgrrangан•type
water AIN J AgrÉ Rex 3*6) 743-64.
104. CD. EC. Wtwn is a rulos_ gun f974;
81
DA in a Ot
1993;Oœ',
- SB, SL SWvm KW _ Cm interfermce ras. 175-82
107. KN_ Ettœts Ot and covor cm rA1rn rnaw
Xuo O , Zhu Z, Stewart BA, DA
X, S, Sim H, poi D, Wau Y_ [hy rmttor, as
water in
110. GP , PV Fritz AK, MB , GA BS. Ot dra.ght and tenwrture
M•at. 2012; t 0711
FPt 124S PMIO: 324807/3
cra Rov PEmtsci_
- 112 Ai N M _ A GrowÜL Qu*y Wat« A
Pflanzen_ 202274:371-U
, A, RoNnson dB, DJ. A tod tor
Erwiron %ftware_ 04 SS | 14. CA.
by J. l): 121—7.
115. , MF. EarWnm JG _ ro»onsa Ot to watx at AgrWatgr Manaw_ https•./doi O. 1
116. A, Kr*ic PN, M•mte AD. Zhau B. SC. MS, al. of a&pt. to in
•meat tarrn to Ag Ecœwt
117. K, A, DI. C, SA, F, Ot al a tinctkmal tor NN
;
PMID 34226560
- | 18. KY. Ot in sssAJ_ 61
VR, R, D, Acoœta-Martinoz V, W of" omrû: io in
4œ:1154W_ 120.
2m1;
121. A, B, AO, UM, OJ- cover wheat•Wbw sy•stxn. J. 20f 9: l (4) 2f l'
SW , SS, Froa-mn MA, %hipanski ME, j, Lal H, Ot _ oxnz• aNo a mat to SSAL 2;
76 (0494-604.
123. SW , IT. J. Cartn•v:
Activa SO' Carbm_ DL, Stott DE. Mikha MM, oeitors. Soil Hoalth Series: Vouyn02
t.xy Sod Hganh 2 p. 152-75.
124. LLEas S. R. Can matter SSSAJ_ 2021
125. S. V. so' Cartxm as an Soit in tho %rtia_ In:
Sahnikov E, Ma_wner LavrÉtEtov A, F , Sod ruw Ya-k:
Cham•, p.

126. Docket H_ Ot cover
M_œ_ (AL): t.
127. JO, Swdt RS, %Gowan JA. CanpariMm Ot

So
u

128. Pan G, Smith P, Pan W. The role of soil organic matter in maintaining the productivity and yield stability of cereals in China. *Agr Ecosyst Environ*. 2009;1; 129(1–3):344–8.
129. Oldfield EE, Bradford MA, Wood SA. Global meta-analysis of the relationship between soil organic matter and crop yields. *Soil*. 2019; 5(1):15–32.
130. Hart MM, Reader RJ. Taxonomic basis for variation in the colonization strategy of arbuscular mycorrhizal fungi. *New Phytol*. 2002; 153(2):335–44.
131. Piotrowski JS, Denich T, Klironomos JN, Graham JM, Rillig MC. The effects of arbuscular mycorrhizas on soil aggregation depend on the interaction between plant and fungal species. *New Phytol*. 2004; 164(2):365–73. <https://doi.org/10.1111/j.1469-8137.2004.01181.x> PMID: 33873560
132. Hart MM, Reader RJ. Do arbuscular mycorrhizal fungi recover from soil disturbance differently? *Trop Ecol*. 2004; 45(1):97–112.
133. Maia LC, Passos JH, Silva JA, Oehl F, Assis DM. Species diversity of Glomeromycota in Brazilian biomes. *Sydowia*. 2020; 72:181–205. <https://doi.org/10.12905/0380.sydowia72-2020-0181>
134. Anderson RC, Libert AE, Dickman LA. Interaction of vascular plants and vesicular-arbuscular mycorrhizal fungi across a soil moisture-nutrient gradient. *Oecologia*. 1984; 64(1):111–7. <https://doi.org/10.1007/BF00377552> PMID: 28311647
135. Deepika S, Kothamasi D. Soil moisture—a regulator of arbuscular mycorrhizal fungal community assembly and symbiotic phosphorus uptake. *Mycorrhiza*. 2015; 25(1):67–75. <https://doi.org/10.1007/s00572-014-0598-1> PMID: 25085217
136. Cavagnaro TR. Soil moisture legacy effects: impacts on soil nutrients, plants and mycorrhizal responsiveness. *Soil Biol Biochem*. 2016; 95:173–9.
137. Cano C, Bago A. Competition and substrate colonization strategies of three polyxenically grown arbuscular mycorrhizal fungi. *Mycologia*. 2017; 97(6):1201–14. <https://doi.org/10.1080/15572536.2006.11832730>.
138. Engelmoer DJ, Behm JE, Toby Kiers E. Intense competition between arbuscular mycorrhizal mutualists in an in vitro root microbiome negatively affects total fungal abundance. *Mol Ecol*. 2014; 23(6):1584–93. <https://doi.org/10.1111/mec.12451> PMID: 24050702